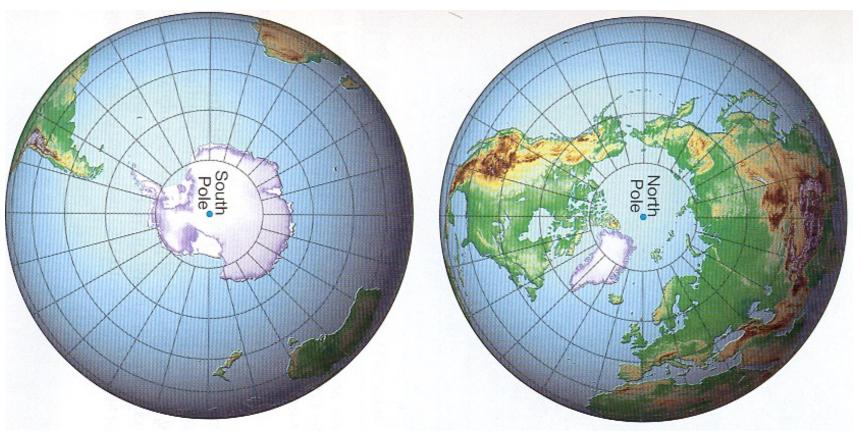
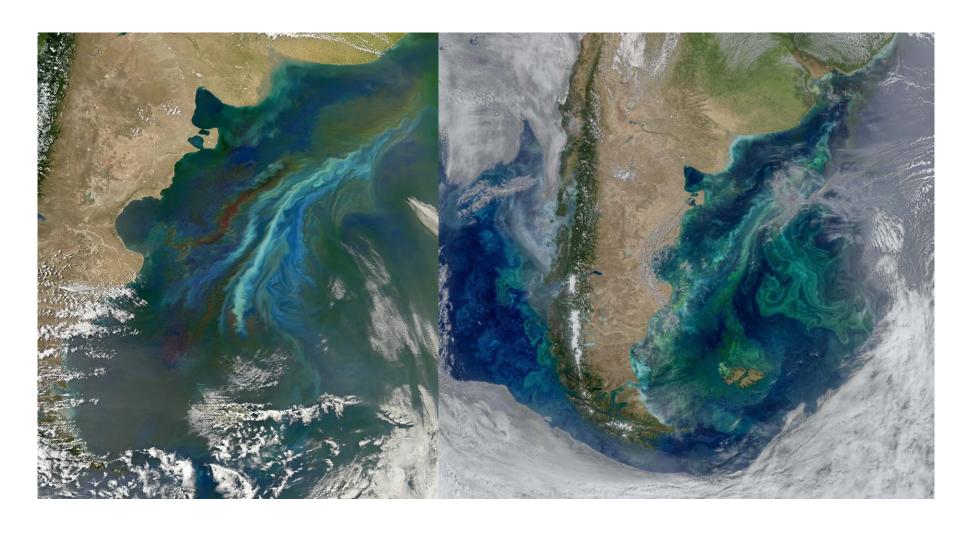
Lab methods & IOCCG Protocols for data collection

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IOCCG Ocean Optics & Biogeochemistry Protocols for Satellite Ocean Colour Sensor Validation

2022 - Fifth IOCCG Summer Lecture Series (18 - 29 July 2022, Villefranche, France)



December 21, 2010 (MODIS-Aqua)

February 4, 2019

How can we improve ocean color information?

Theory and fundaments

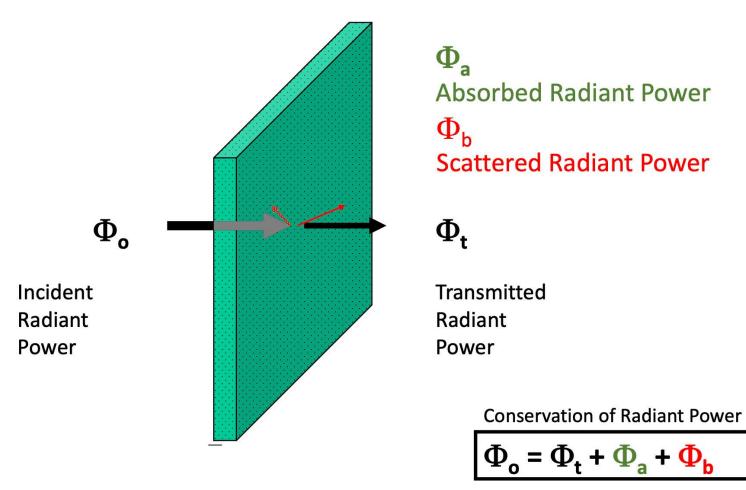


As light penetrates the ocean surface and propagates to depth, what processes affect the light transfer?

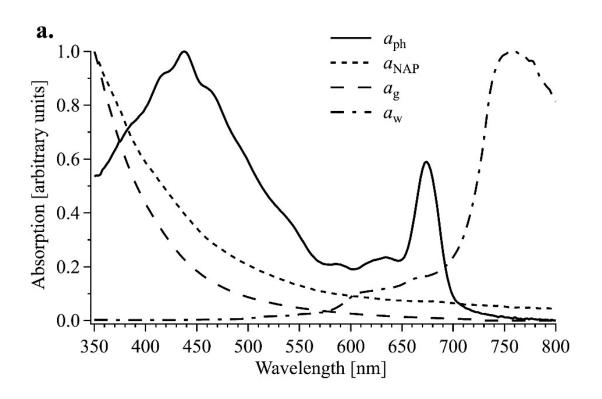
- Absorption removes light
- Scattering redirects light
- Re-emission converts from one wavelength to another (one direction to another)
- A portion leaves the surface



Consider loss due to *beam* attenuation (absorption + scattering)

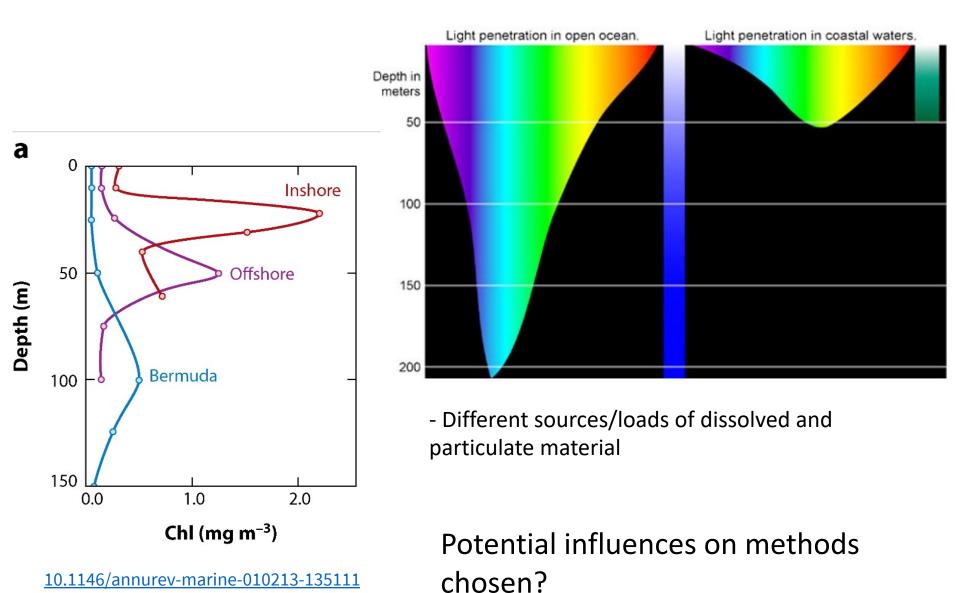


Spectral light absorption and its components

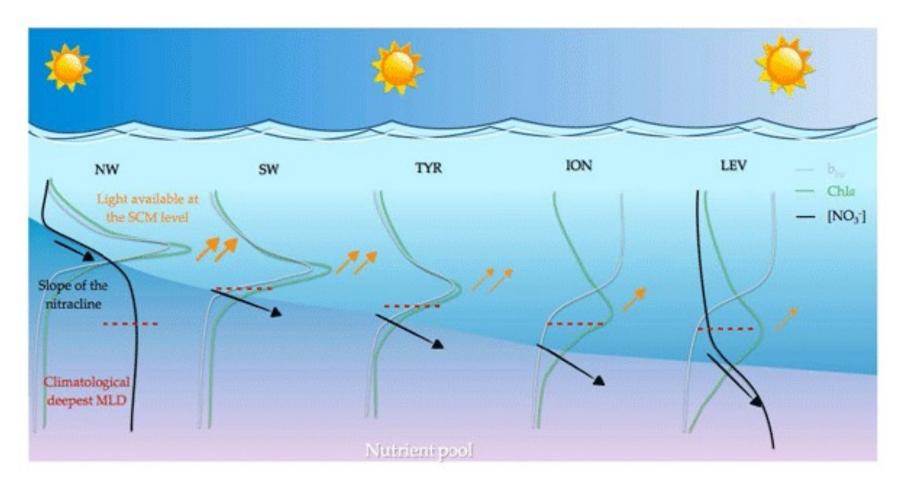


Why do you want to measure it?
What do the components represent?
How can you measure it?

Where do you want to measured it?



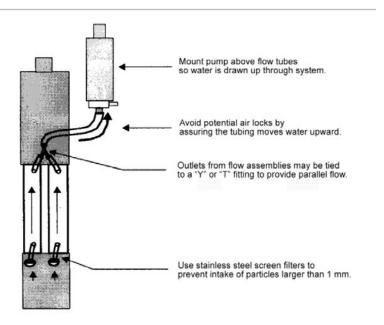
Seasonal and spatial gradients (horizontal and/or vertical)



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Potential influences on methods

Field measurements



Profiles, Moorings
In line - Pump

Corrections for T, S and scattering Bubbles, Non algal particles, CDOM can be measured with a capsule filter, help to correct for scattering as well

- Integrating Cavity Absorption Meters
- Point-Source Integrating Cavity Absorption Meter

Discrete samples: filter pad – glass fiber

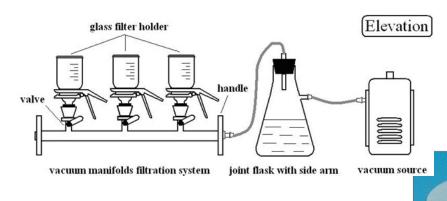
- Standard procedures – standard brands



GF/F nominal pore size 0.7 microns

Highly scattering – differences among boxes/lots





Some good practices for sample collection and handling

- Common errors
 - Settling of particles standard gentle mixing
 - Filtration volumes ideally optical density should be around 0.1 to 0.4 – modify filtered volumes
 - Time to process samples immediate scanning or freezing and storage – use liquid nitrogen
 - Forget to store blank filters (filtered with milli-q)with samples
 - Monitor vacuum pressure should not exceed 5-10 mmHg or 0.1-0.2 psi – spots
 - Gelatinous organisms and colonies in the central area of the filter

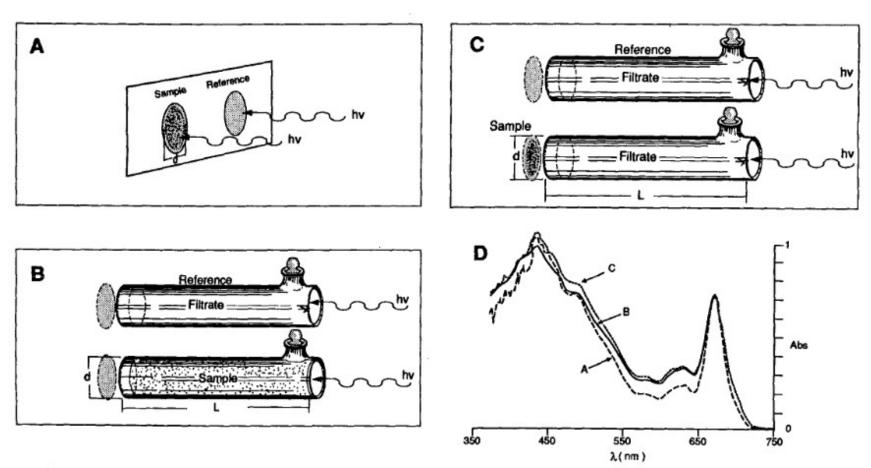
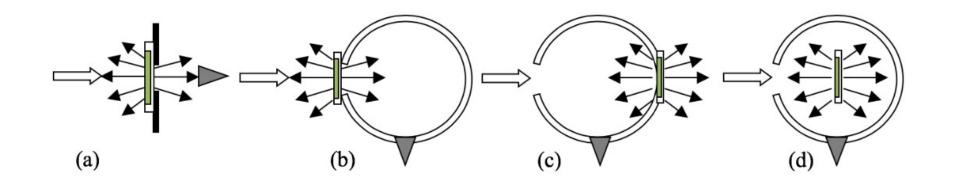


Fig. 2. Cuvette calibration for the filter technique. A. Filter technique used at sea. B. Cells in a 10-cm cuvette, 1.9-cm i.d., with blank filters (GFF) for diffusers. C. Cells on filter at exit of cuvette; blank filter is diffuser reference. D. Absorption spectra of panels A-C for *Phaeodactylum tricornutum*. Filtered media used in reference cuvette.

What kind of Equipment do you have access to for the measurements?



- a) transmittance mode (T-mode),
- b) transmittance and c) reflectance mode measured with an integrating sphere with externally mounted samples (T-R method); (d) internally mounted sample in integrating sphere (IS-mode).

UV-Vis de doble haz real, marca Peaks Instruments. modelo C 7200 S







T-R method

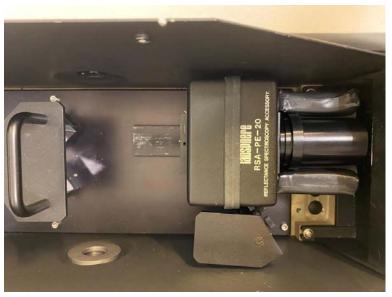
Baselines, drift and lamps





• Pathlength Amplification Correction





Some good practices for sample measurements

- Notch the edge of each filter to provide a means for identifying the orientation in the spectrophotometer
 - pigment extraction
 - Methanol not effective for water soluble
 - NaOCl (0.5-1%)- oxidizes shifts in UV
- Keep the filters moist and protected from light between measurements
- Keep filter orientation
- Monitor stability of the spectrophotometer
- -air measurements with time (30 60 min) but changes with lamp use
- Field and lab notes / protocols as "live documents"
- can anyone (including me :) repeat what I did with my notes?

Computations

$$a(\lambda) = \ln(10) OD(\lambda)/L$$

$$L = (V/A)$$

$$a_x(\lambda) = \ln(10) \ OD_s(\lambda) / (V/A)$$

T-mode:
$$OD_s = 0.679 (OD_f)^{1.2804}$$
 (5.5)

T-R-mode:
$$OD_s = 0.719 (OD_f)^{1.2287}$$
 (5.6)

IS-mode:
$$OD_s = 0.323 (OD_f)^{1.0867}$$
 (5.7)

