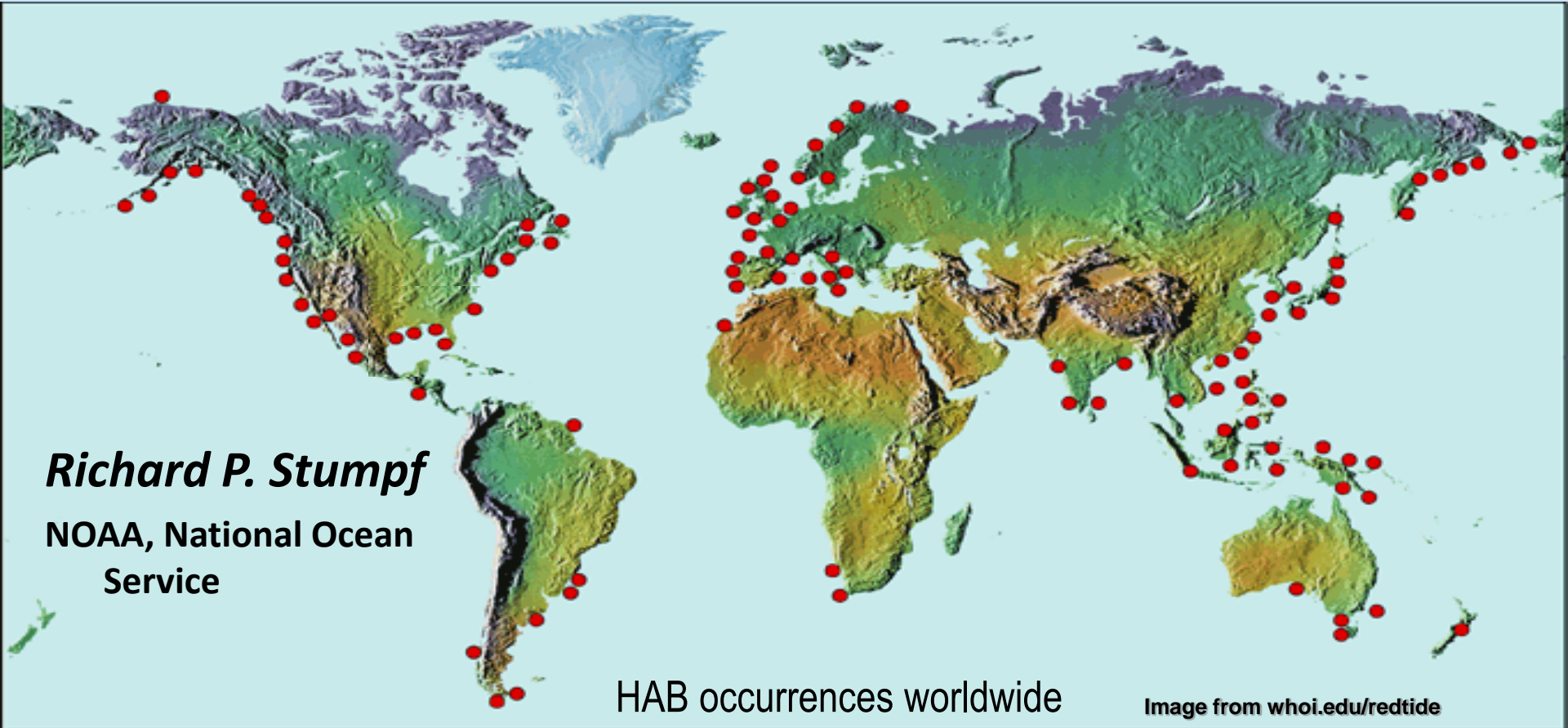


Harmful Algal Blooms (HABs) 2 methods



Remote Sensing for HAB Detection: Does the Bloom Change Water Color?

Yes? Optical techniques

- Can distinguish some bloom types in case 1 water

- Difficult in coastal areas due to sediment and CDOM

- Use absorption, backscatter, and spectral shape, relative patterns

- Complements ecological detection techniques

- Many blooms are not unique (Pn is a diatom in a diatom system)

Bloom Ecology techniques, **Not just chl *a***

- Does it dominate/correlate biomass?

- Seasonality of blooms

- Relationship to environmental conditions

No? Physical Forcing

- Association between bloom and physical features, **use of SST**
(e.g., fronts, upwelling, wind events)

- Not direct bloom detection, difficult to view extent of bloom

***Karenia* can make strong blooms (major source of primary production on west Florida shelf). Pure *Karenia* blooms are great for Ocean Color Remote Sensing, gave illusion that “red tides” are easily solvable.**



Credit: Paul Schmidt

**If you want a “red tide”: *Nocticula*.
Visually striking; usually harmless, sometimes causes fish kills**



Cyanos in many areas

Baltic

Large lakes (e.g. Lake Erie)

Small lakes



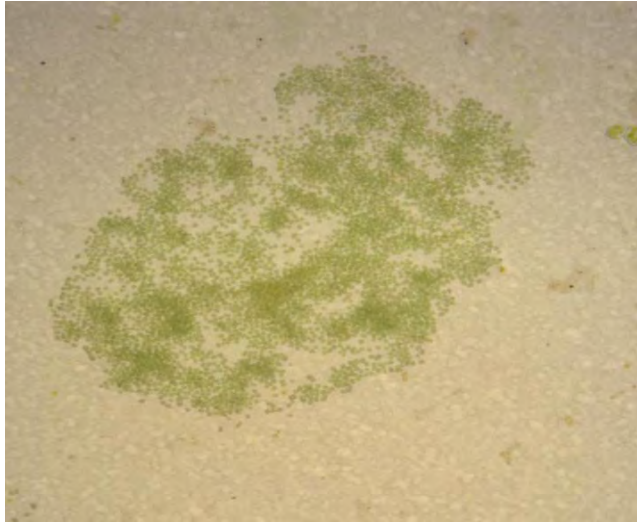
MERIS 08-Oct-2011

100 km

This satellite image from MERIS on October 8, 2011, shows a cyanobacterial bloom in the Baltic Sea. The bloom appears as a large, irregular green area extending from the coast. A 100 km scale bar is located at the bottom right of the image.

***Microcystis* as scum example**

- Aug 2009, Lake Erie



Remote Sensing & Optical methods for detecting and monitoring HABs

Colored blooms

Uniquely identify bloom by Optical techniques

Identify HAB

Identify non-HAB

Identify blooms through relationships

dominate biomass?

Ecological/environmental associations?

Non-Colored blooms

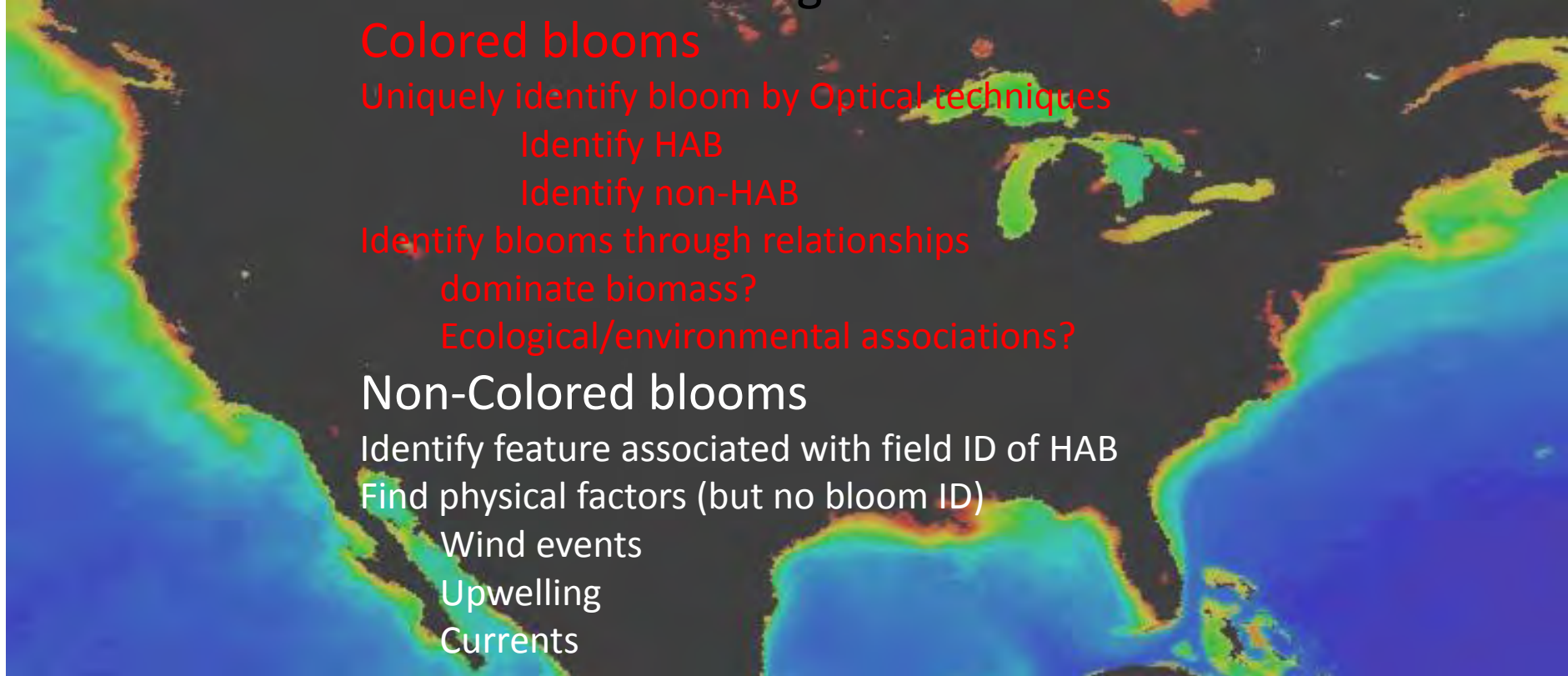
Identify feature associated with field ID of HAB

Find physical factors (but no bloom ID)

Wind events

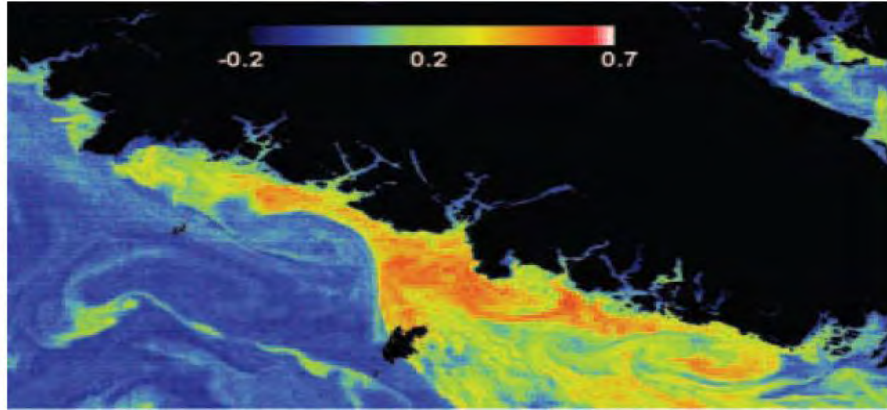
Upwelling

Currents



Chlorophyll

Generally not useful simply as chlorophyll
(Chlorophyll may be inferred from FLH)

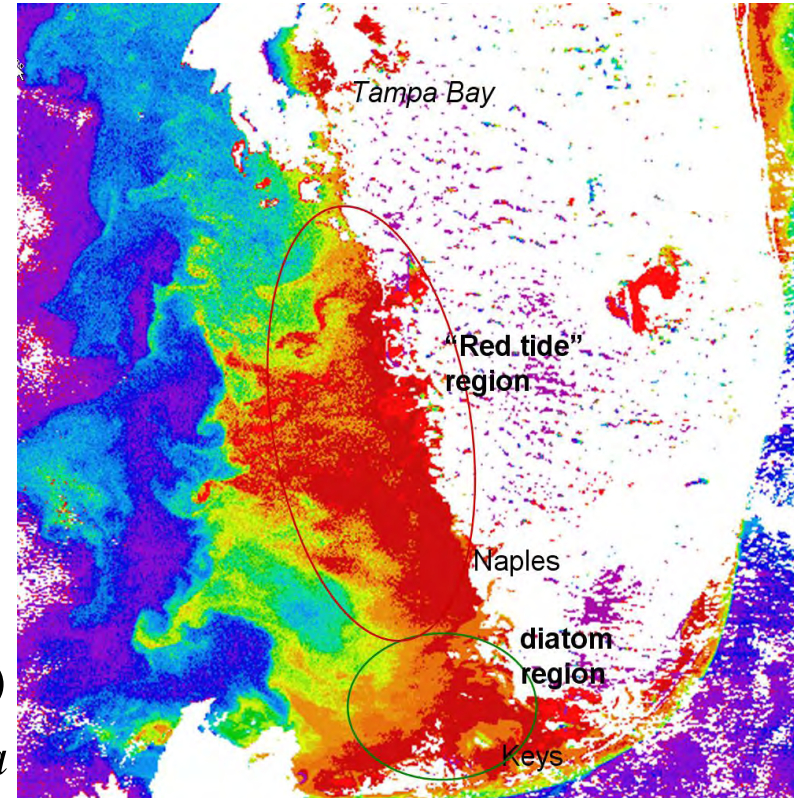


Gower et al., 2005, MERIS FLH

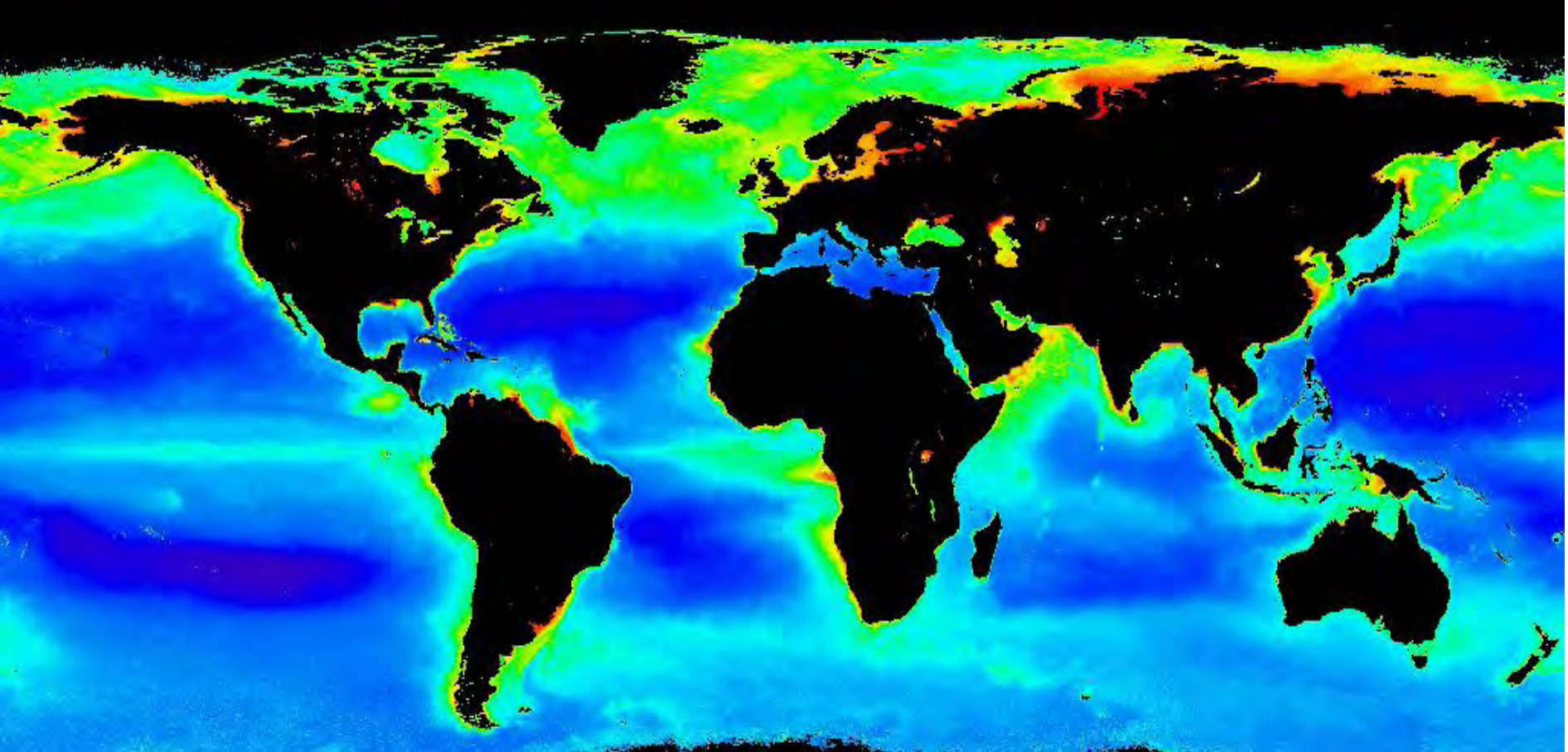
Exceptions, chlorophyll threshold is useful:

“red tide” in some bays in Japan (various species)

Brown tide in US, *Aureococcus*, *Aureoumbra*



Why not chl-a?

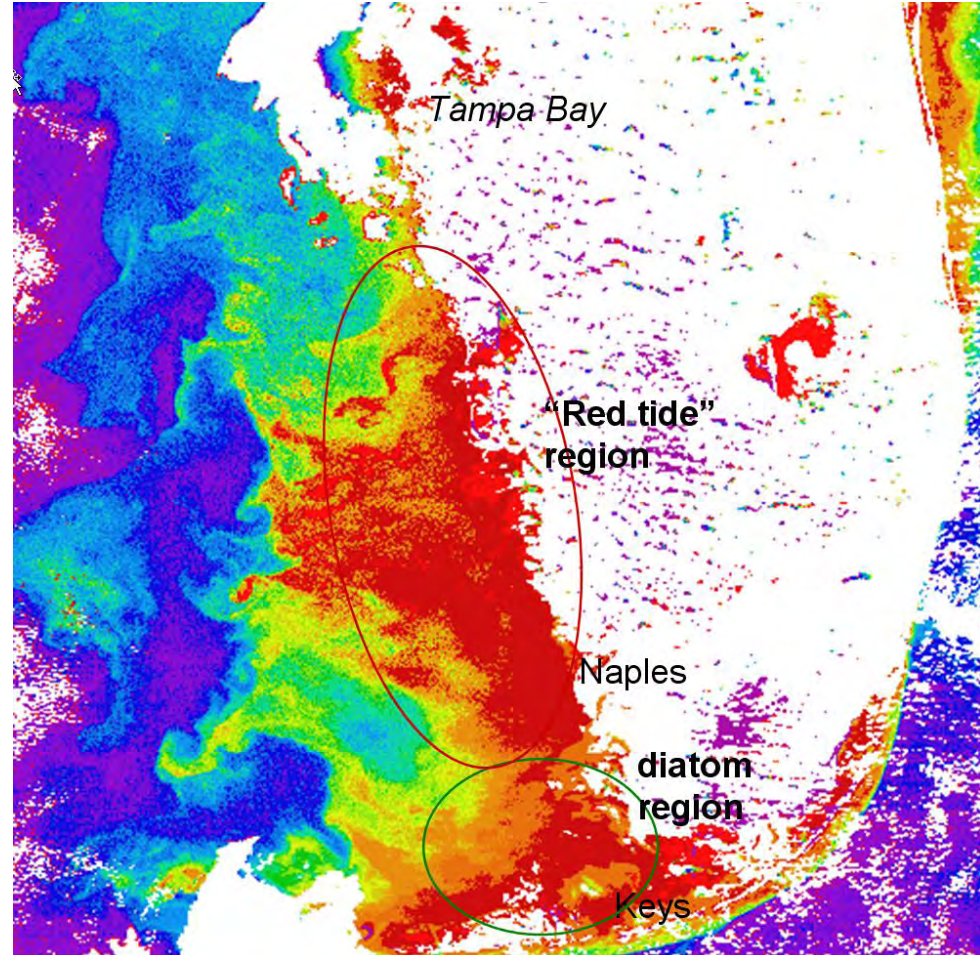


Chlorophyll is usually used in an anomaly or “delta” technique

Qualitative anomaly.

This area is a “red tide” because we infer concentration is much higher than normal.

Most “chlorophyll” HABs are interpreted anomalies



Anomaly method can be quantified

Anomaly finds *New Blooms*

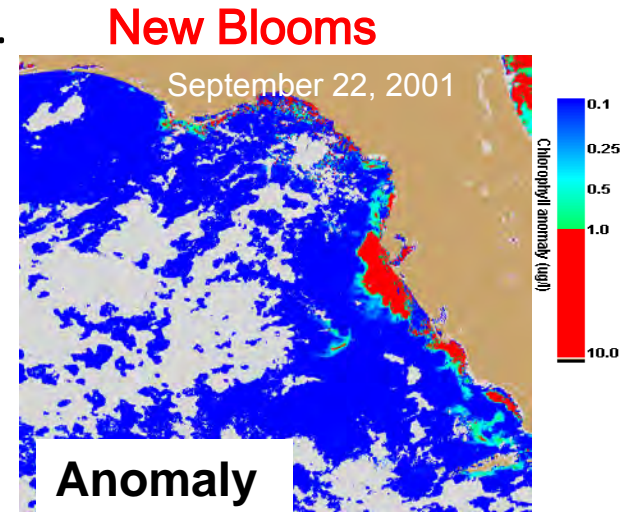
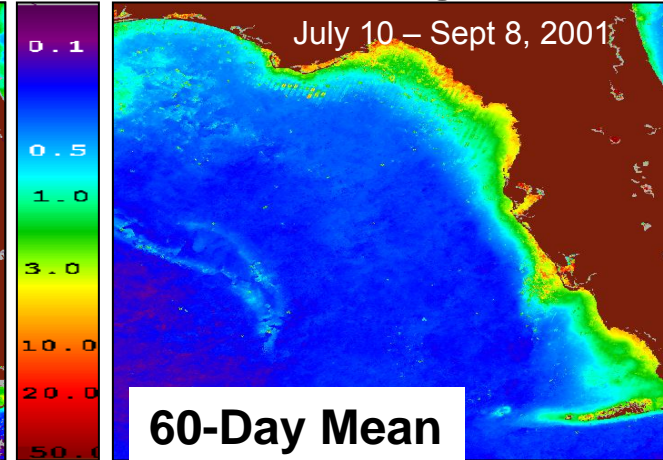
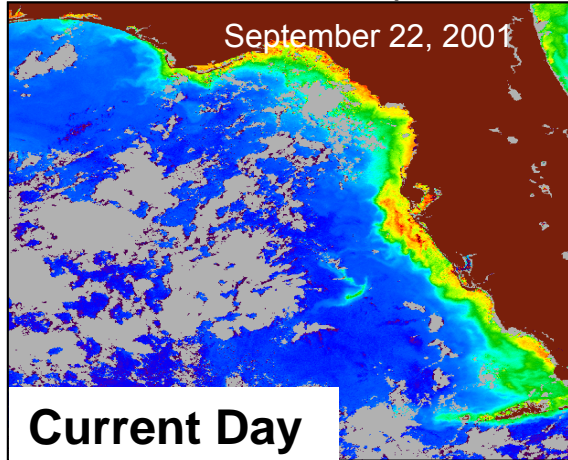
Demonstrated as effective for *Karenia brevis*

K. brevis dominates biomass in late summer

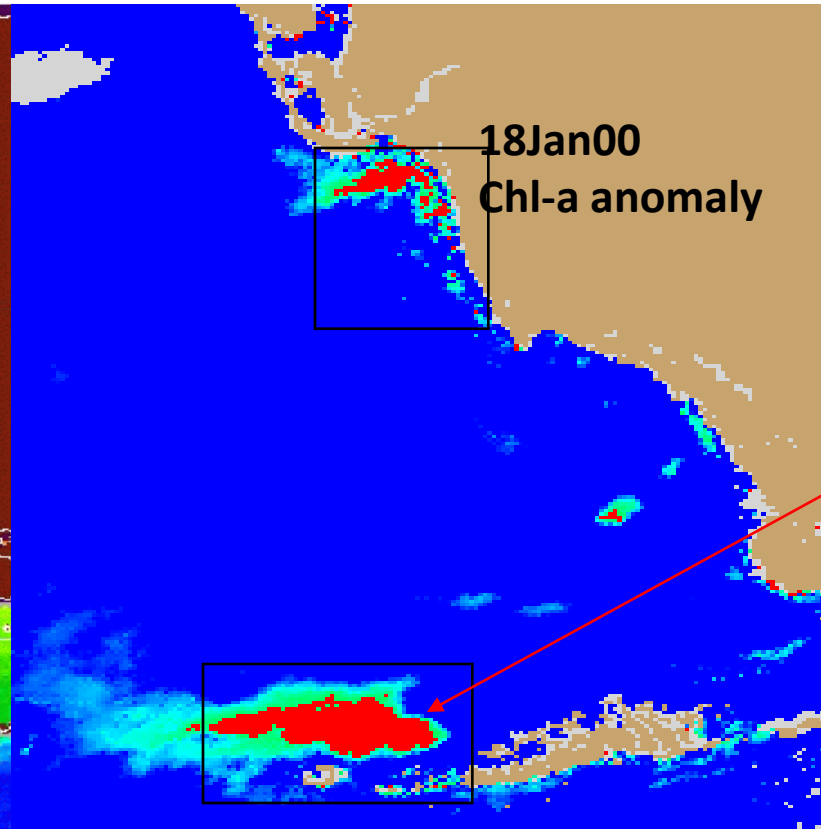
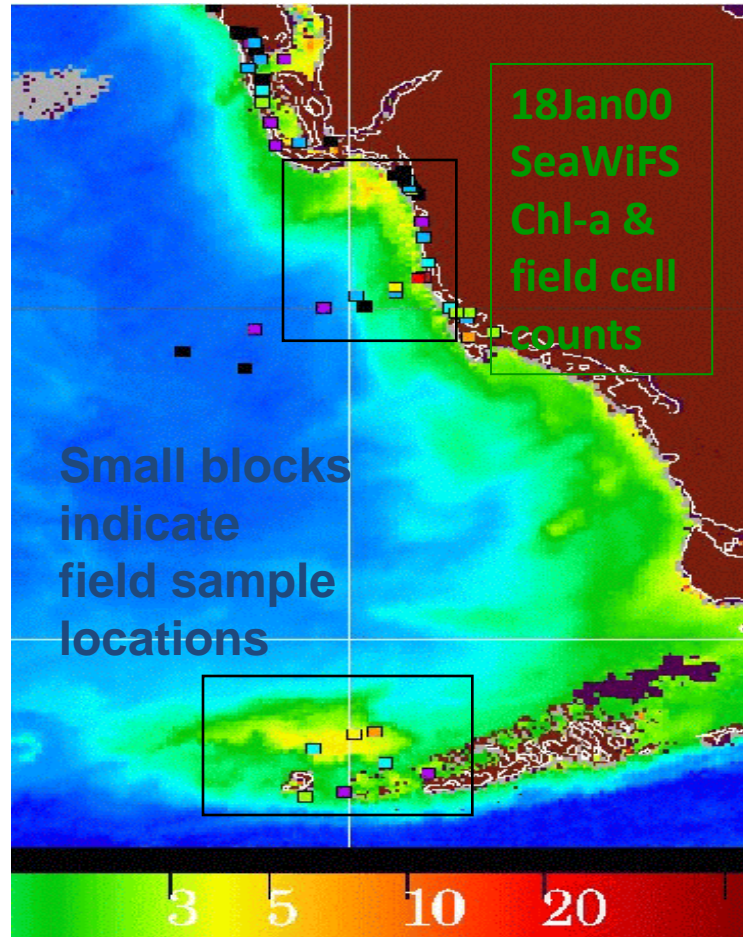
Better in case 2 water (with turbidity) than optical algorithms

Accuracy > 80% during summer and fall

False positives common, False negatives rare.



HAB Detection from SeaWiFS

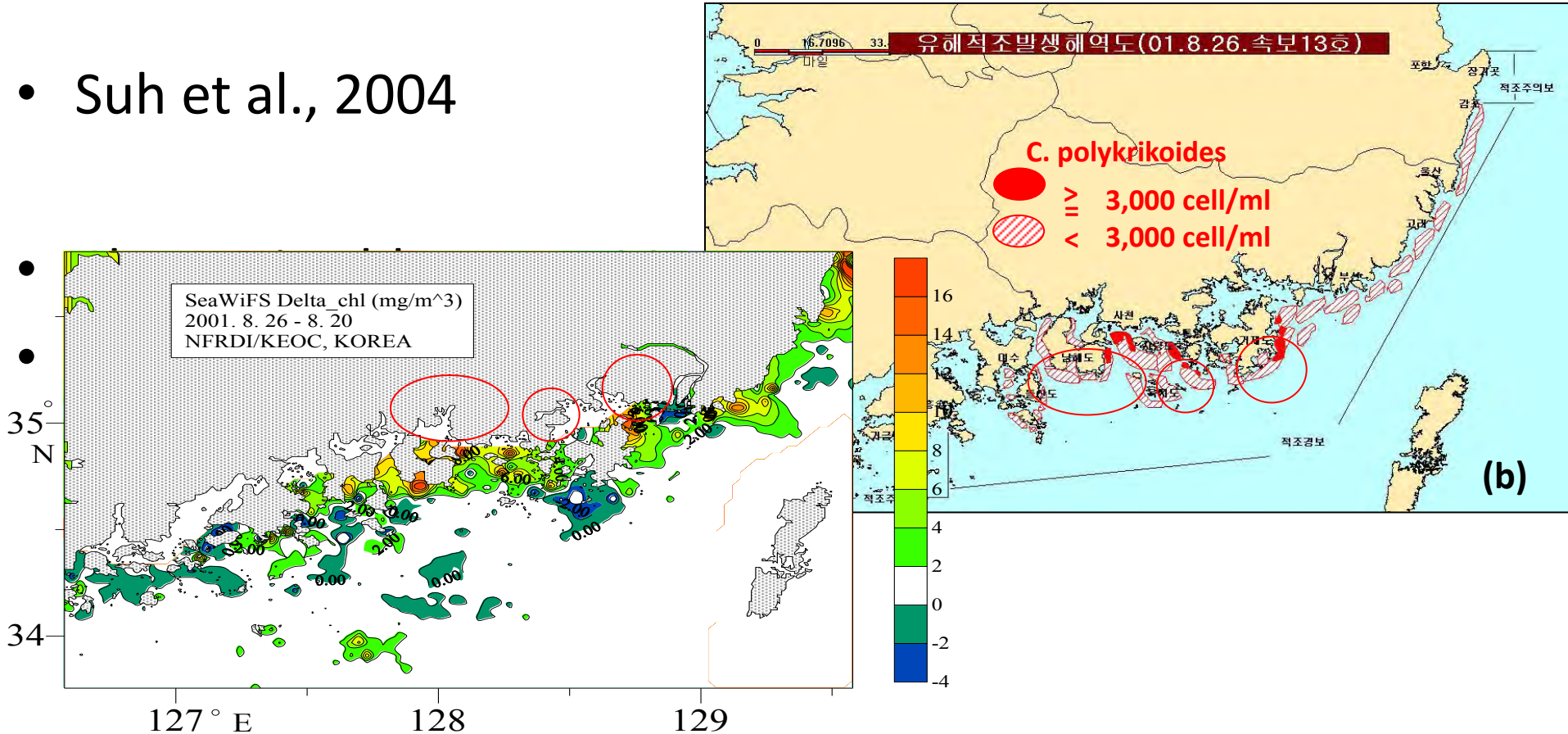


**First HAB
detected
from
satellite
prior to field**

- Bloom extent not identified by field sampling, but satellite provides full size

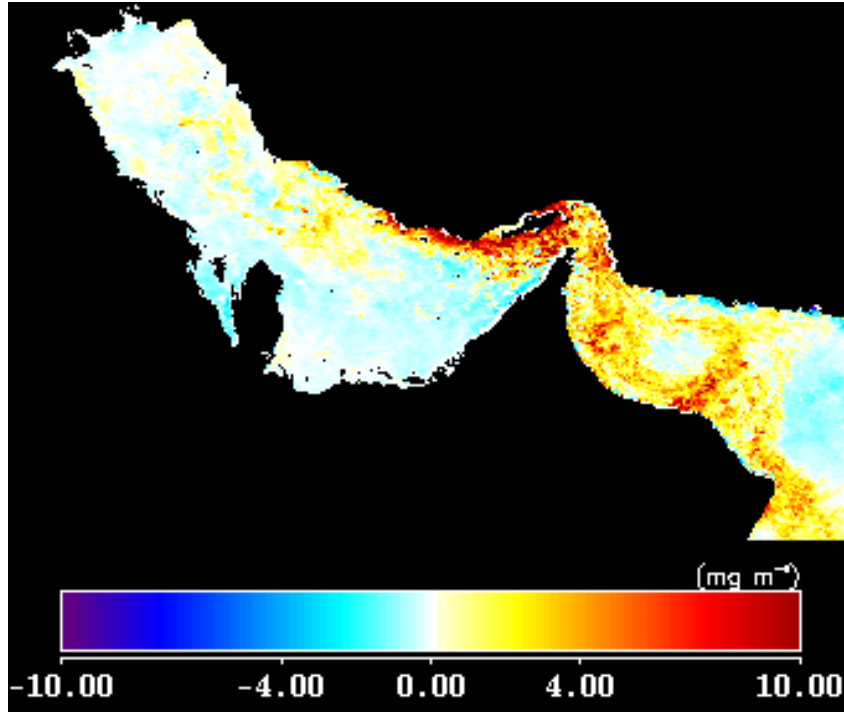
Change in chlorophyll, example for Korea

- Suh et al., 2004

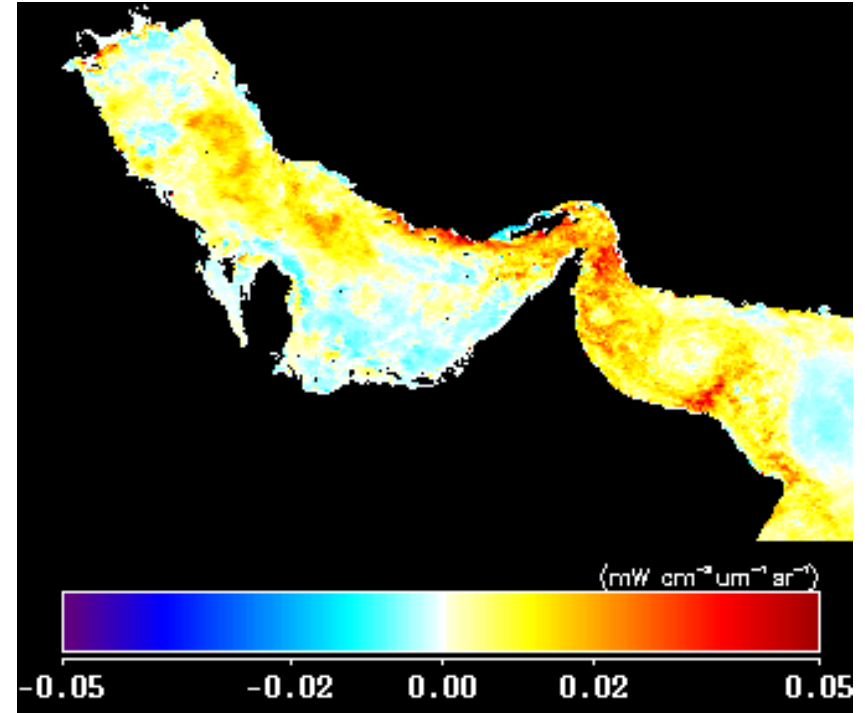


Persian Gulf during *Cochlodinium* bloom

MODIS Chlorophyll anomaly, Nov 2008

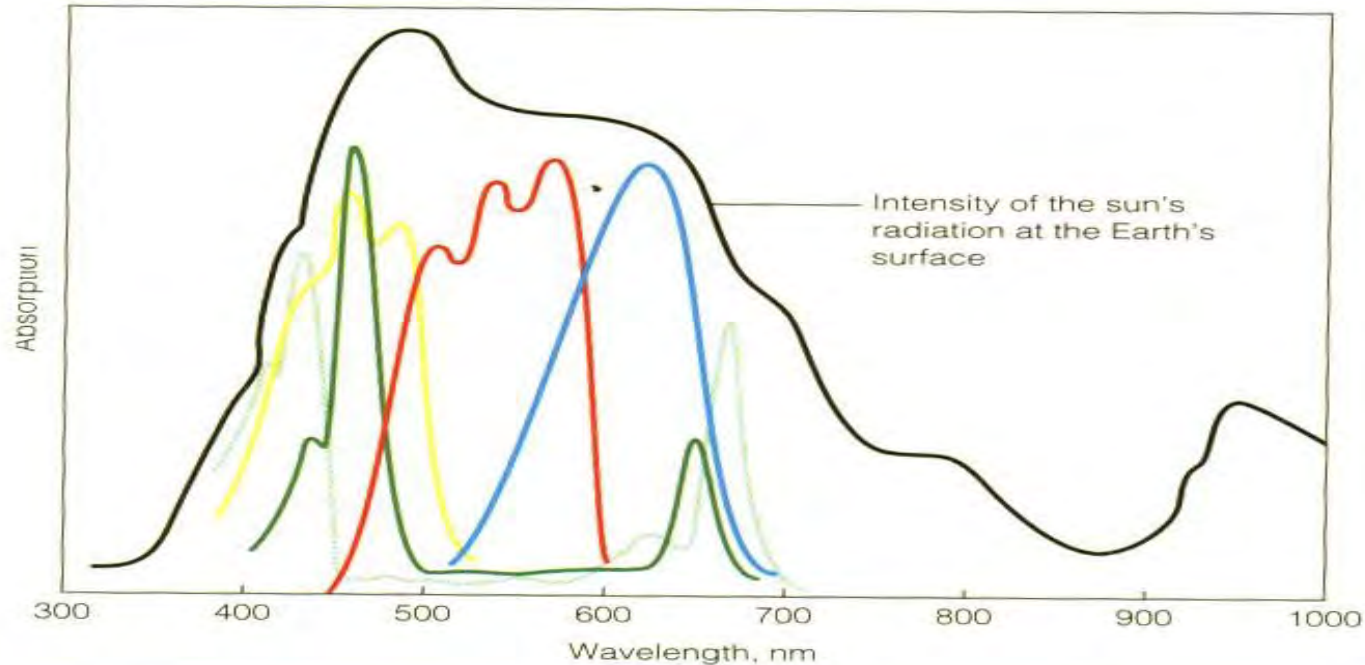


FLH anomaly



From: R. Kudela UCSC

Spectra: various pigments in algae



Key:

— Chlorophyll <i>a</i> (green)	— Phycoerythrin (red)
— Chlorophyll <i>b</i> (green)	— Phycocyanin (blue)
— β carotene (yellow)	

S. Berg Winona State

http://course1.winona.edu/sberg/Fac_sb.htm

Analytical algorithms

- Phycocyanin, from Simis, Gons and others
- QAA for absorption (Lee and others)
 - Solve for absorption at MODIS or SeaWiFS bands
- With good data, quite effective; demonstrated to work with radiometry; satellite depends on atmospheric correction

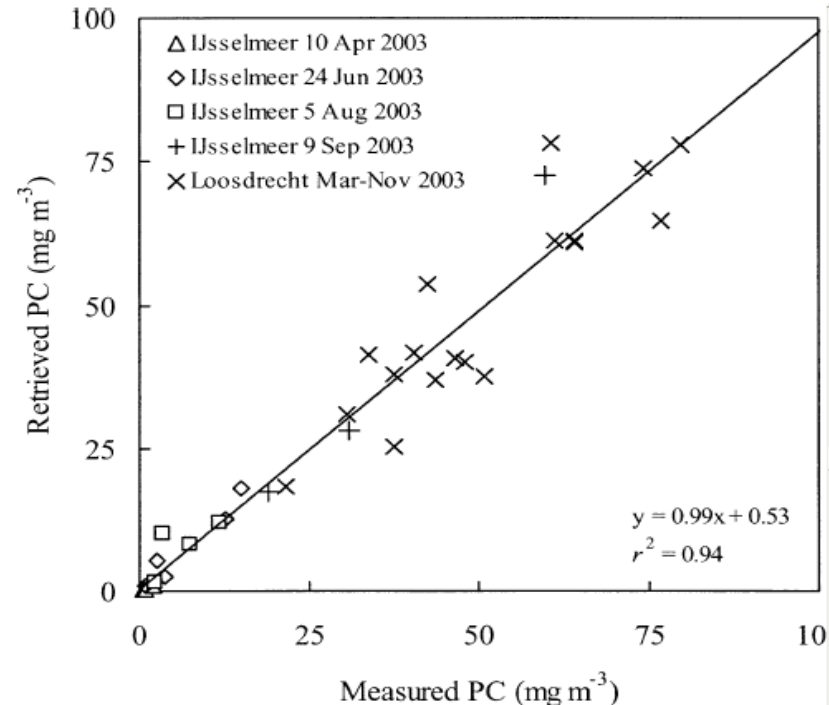
Analytical optics, example from cyanobacteria

- Simis et al. (L&O, 2005) Netherlands, MERIS

Solve for $a_{pc}(620)$ from
 $R(709)/R(620)$

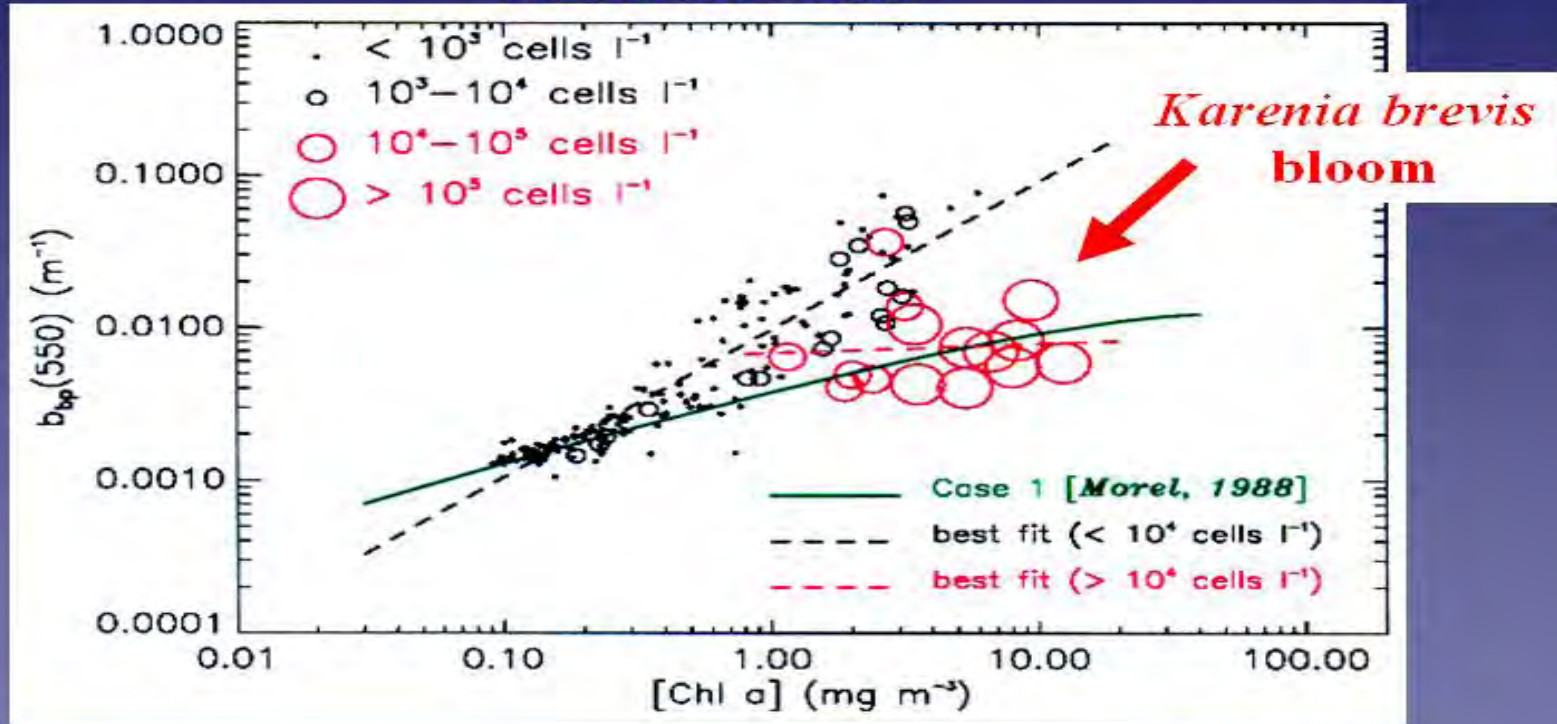
With b_b , a_w , a_{phi}

$$[PC] = a(620)_{pc} / a_{pc}^*(620)$$



Empirical, backscatter against absorption, example from *Karenia*, less scatter from cells and from cellular detritus

West Florida Shelf



Cannizzaro et al. 2002

Spectral shape algorithms

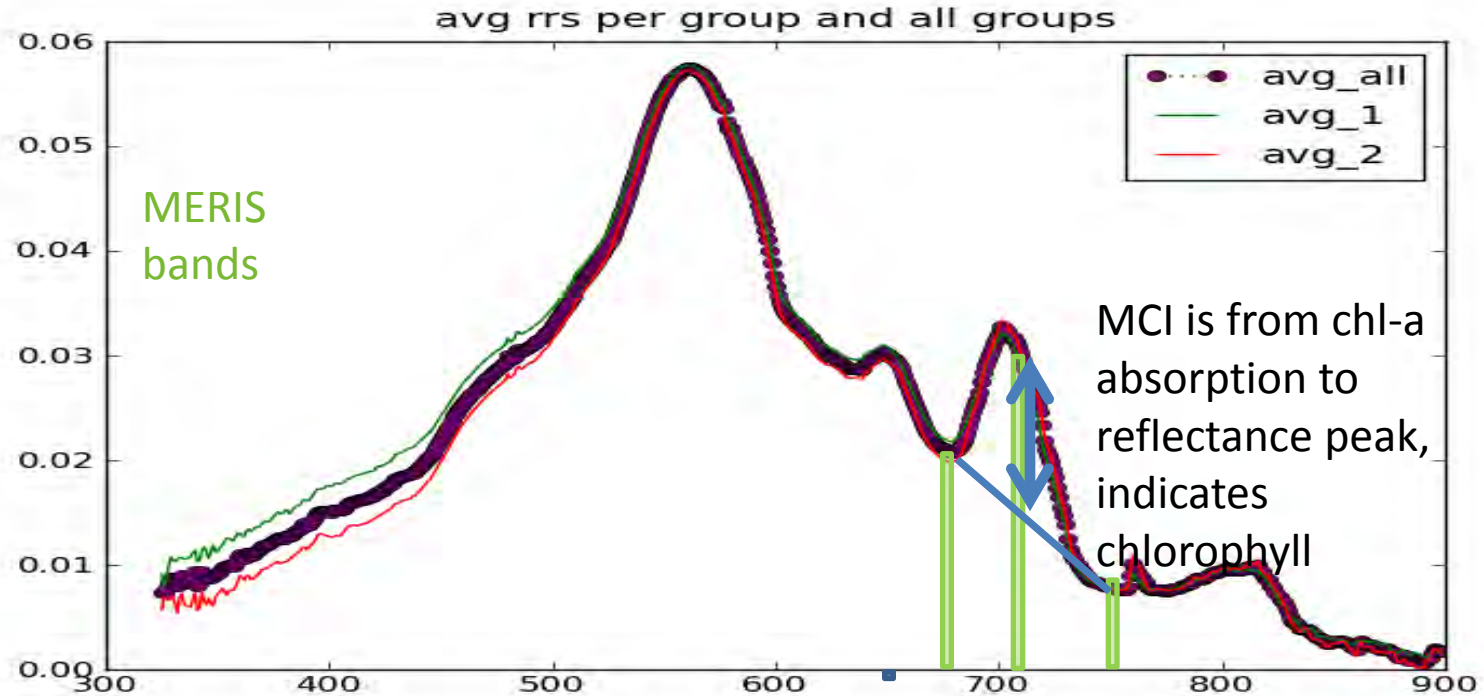
MCI on MERIS (Gower 2005); (quantify by Binding 2011)

CI on MERIS (Wynne et al., 2010)

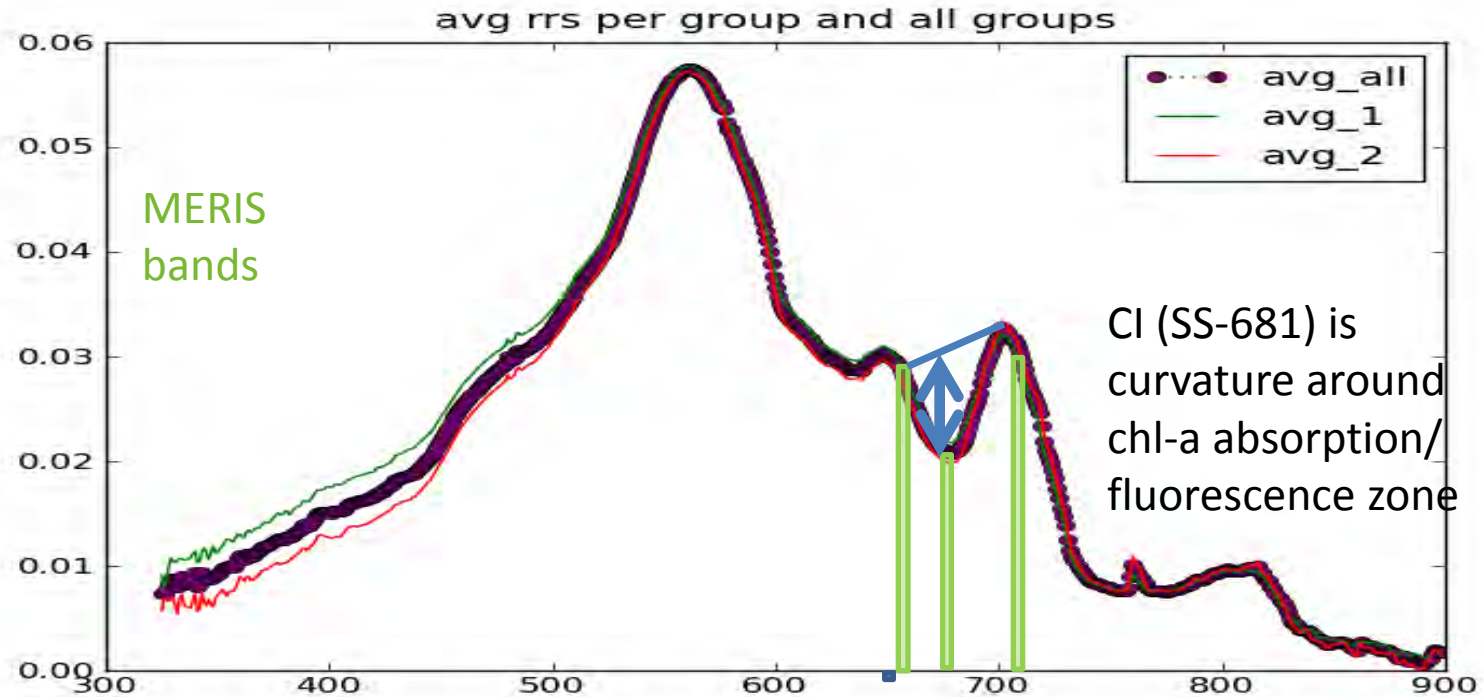
FLH MODIS (applied to a HAB, Hu et al., 2008)

Other shape algorithms (SeaWiFS or MODIS; Tomlinson et al., 2010)

MCI determined at red-edge, related to chlorophyll



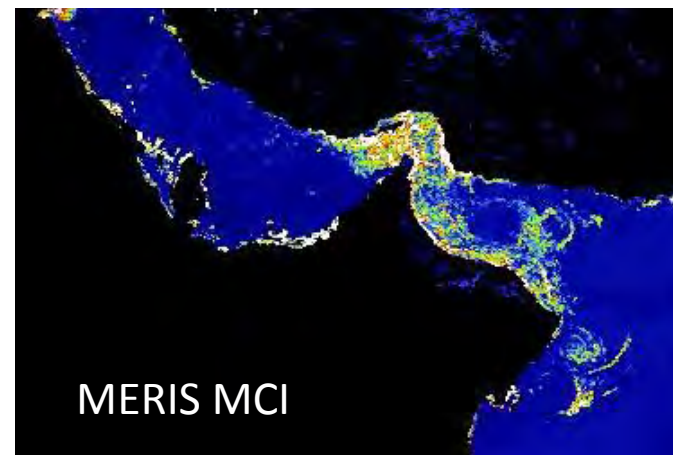
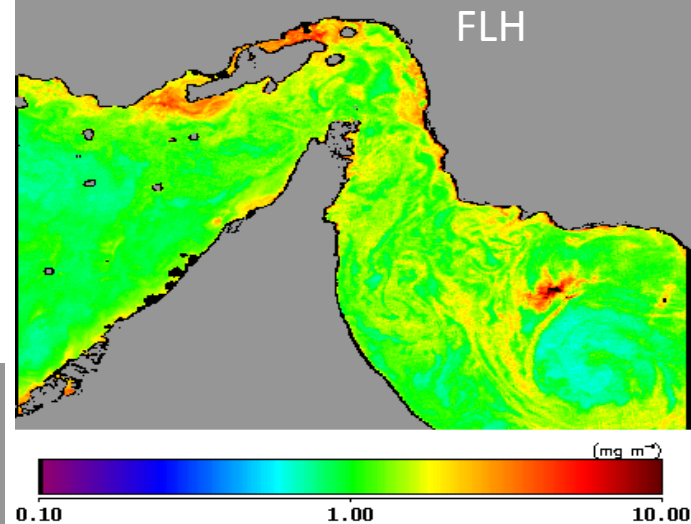
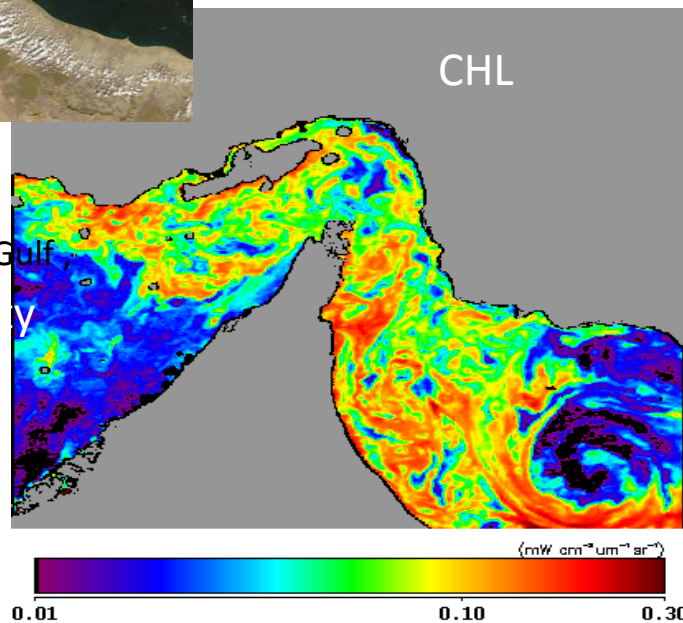
CI/SS681, intense blooms, more cyano sensitive





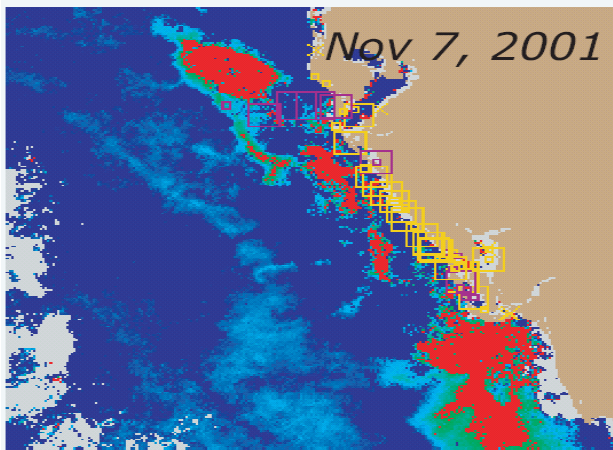
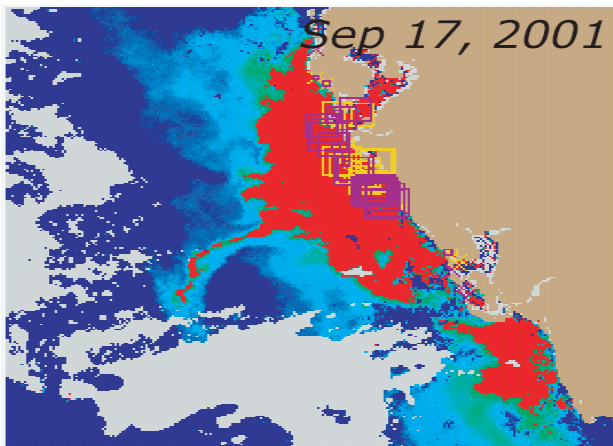
MODIS 19 November 2008;
Cocchlostinium bloom, Persian Gulf,
impacting water supply .

chlorophyll, FLH.
Figures from Raphe Kudela

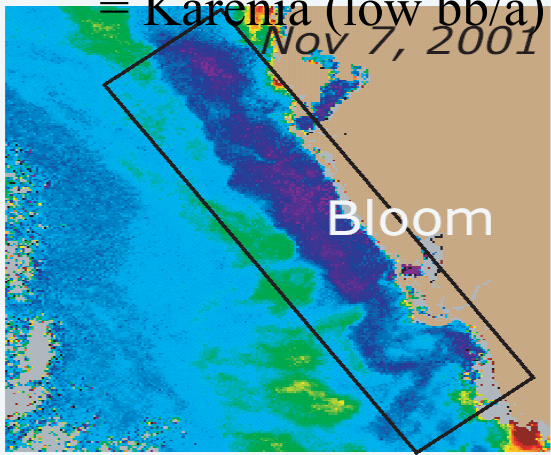
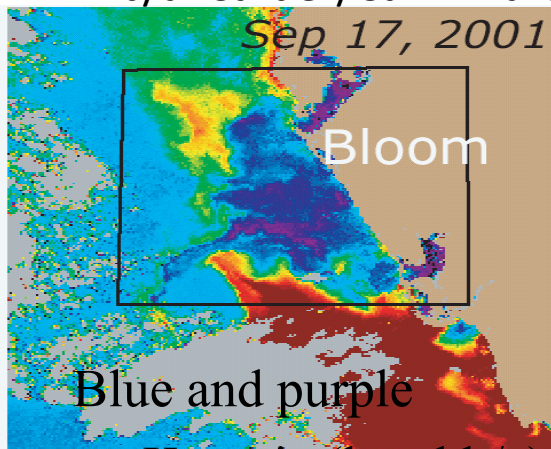


Other Optical Algorithms

Chl Anomaly



Bb/a Carder/Cannizzaro



Bb/a
(Cannizzaro/Carder)

Karenia blooms scatter less than diatom & *Trichodesmium*.

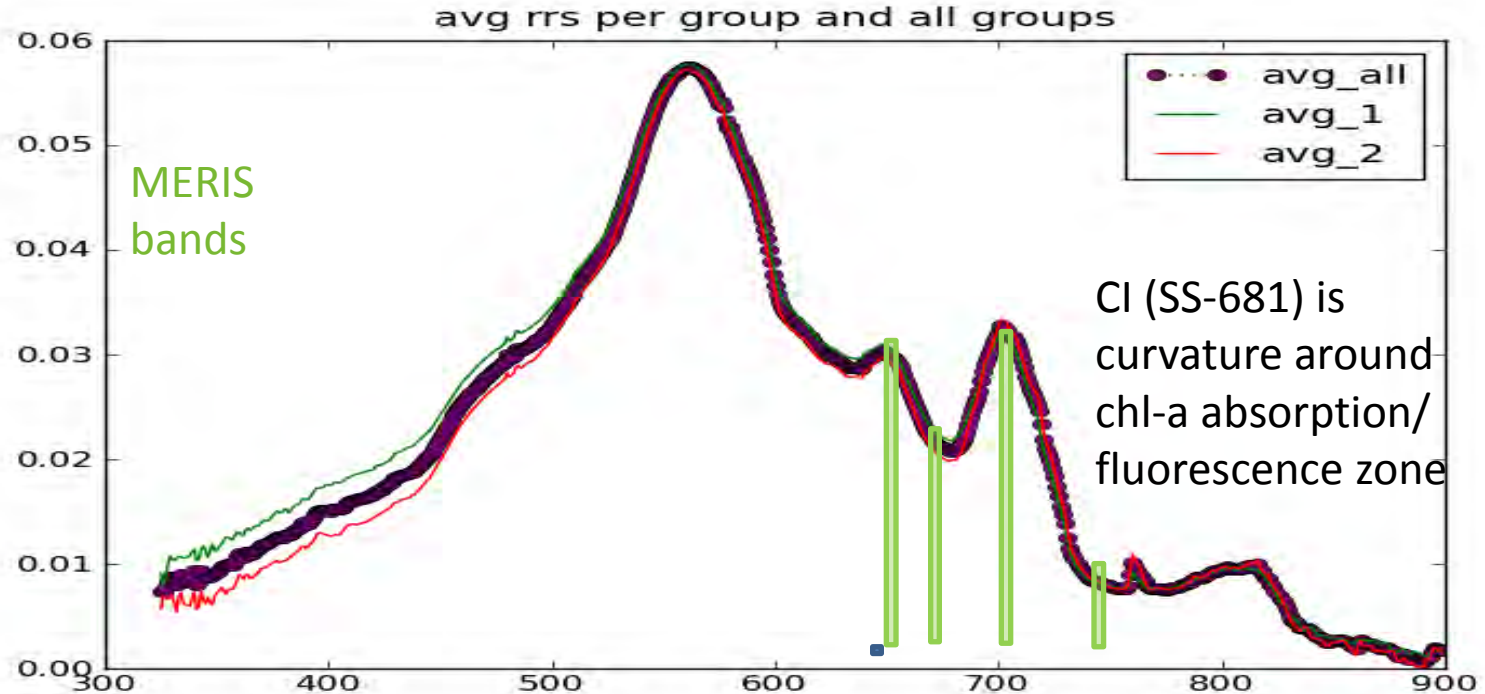
Use Morel bb for reference. *Karenia* bloom when

$(bb/a) < \text{Morel } bb/a$.

bb from Rrs or Qaa, chl or FLH as surrogate for a.

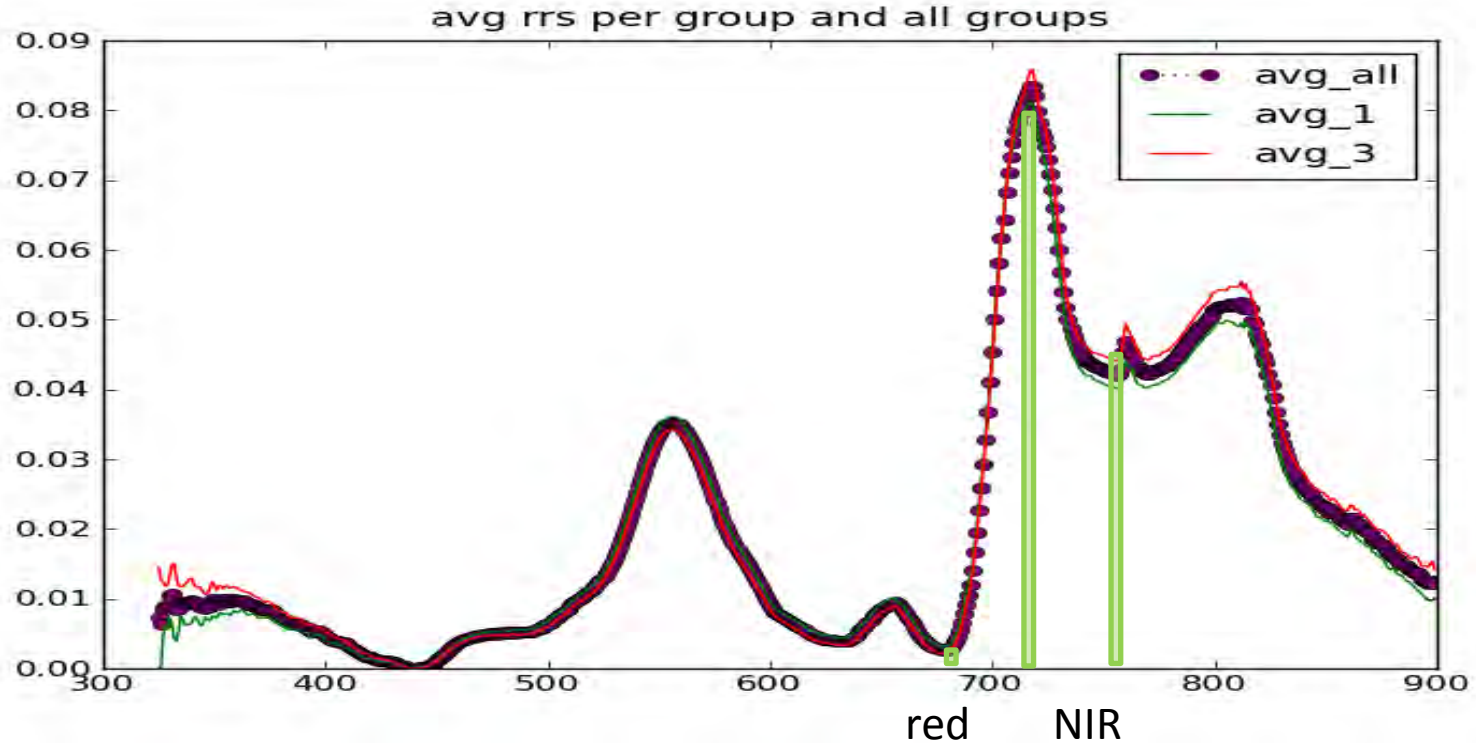
Low is *Karenia* bloom.

intense blooms in water

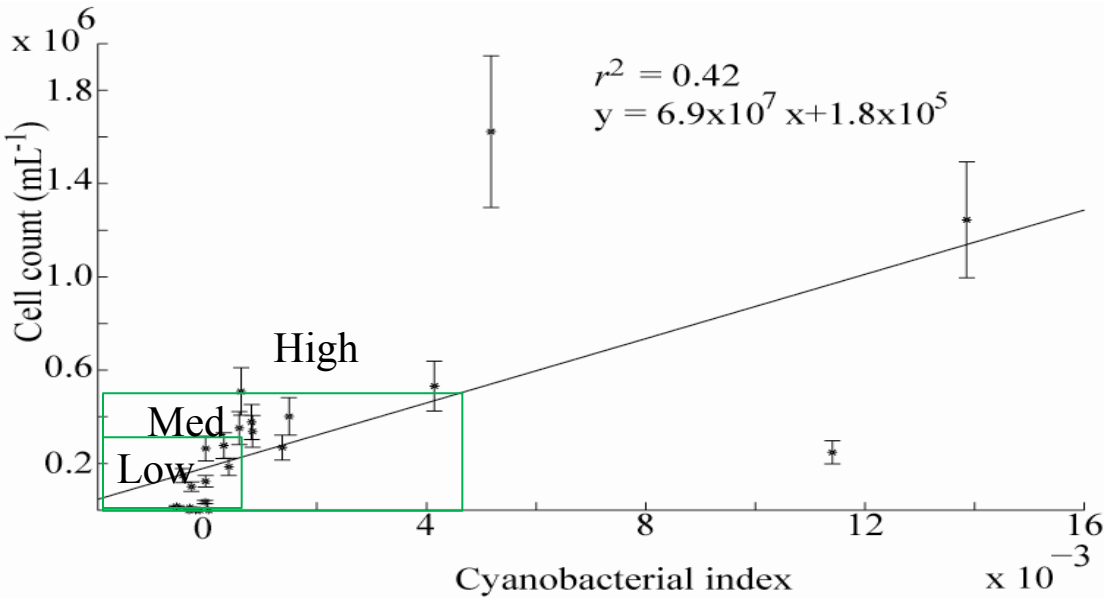


Spectra of *Microcystis* “scum”, bloom on water

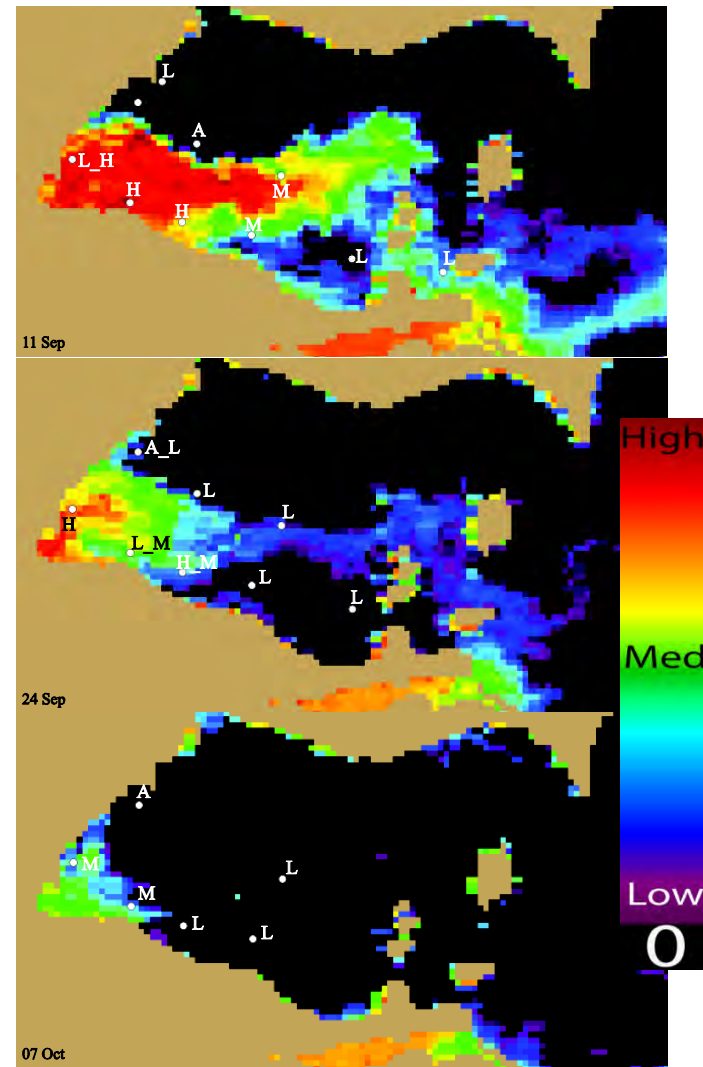
High in NIR, low in red



Obtaining cell counts from satellite

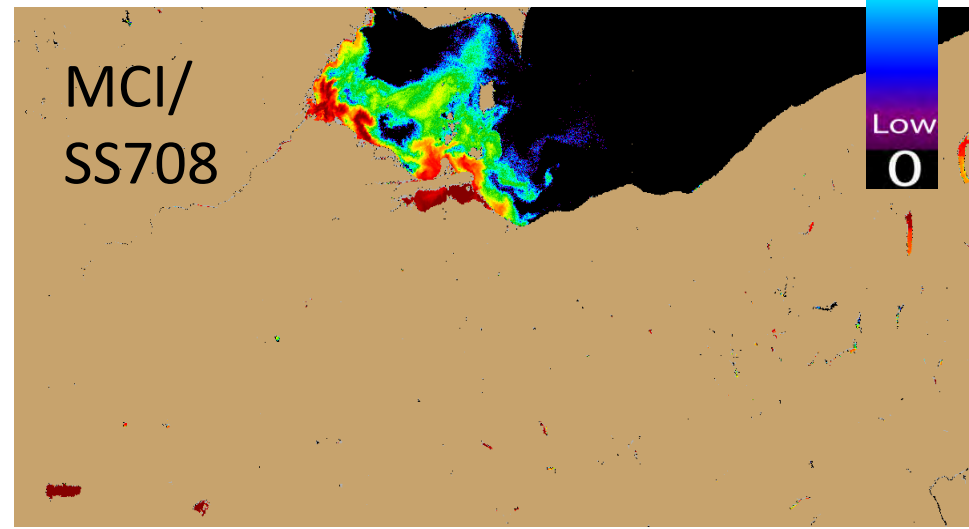
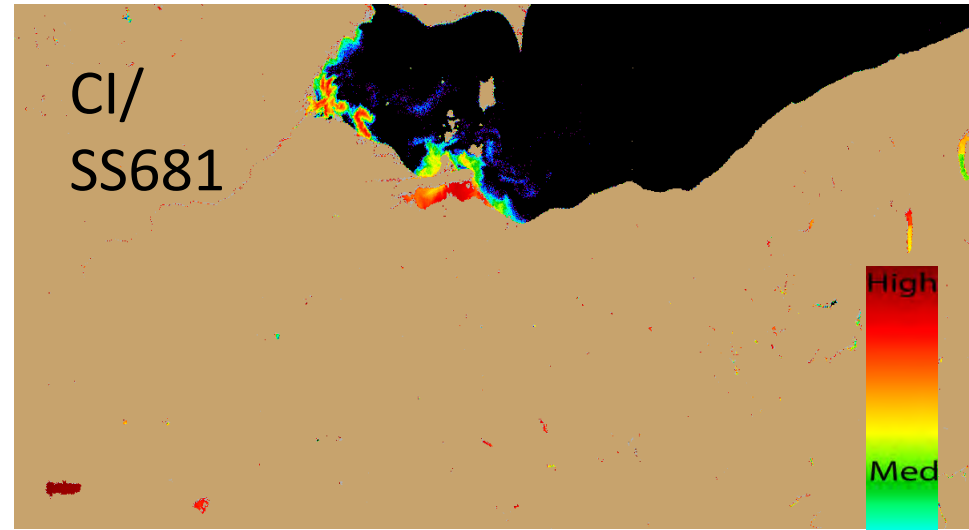


Satellite against field cell counts (now working on field radiometry)



MERIS spectral shapes, cyano & other bloom

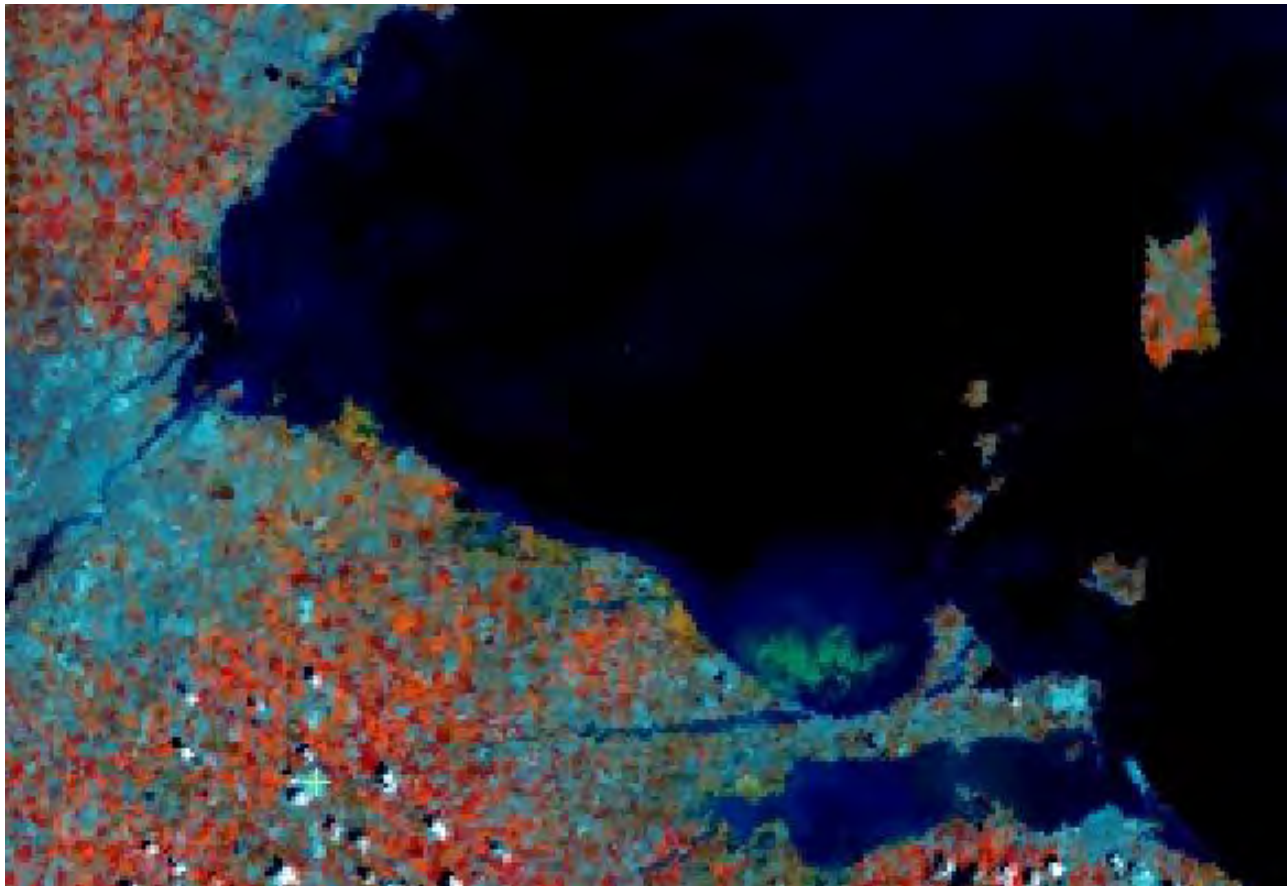
MCI sensitive to lower chl-
a, but also sensitive to high
sediment concentration



**A side note, true color is valuable but hard to
interpret: Find the scum, July 27, 2010**



NIR and red



Other sensors

Cyanobacteria under high concentrations and low winds produce scum, high NIR reflectance

NIR assigned red color

*False-color IR Landsat,
Potomac River, 1982*



***Anabaena* bloom, AVHRR (only red and NIR)**

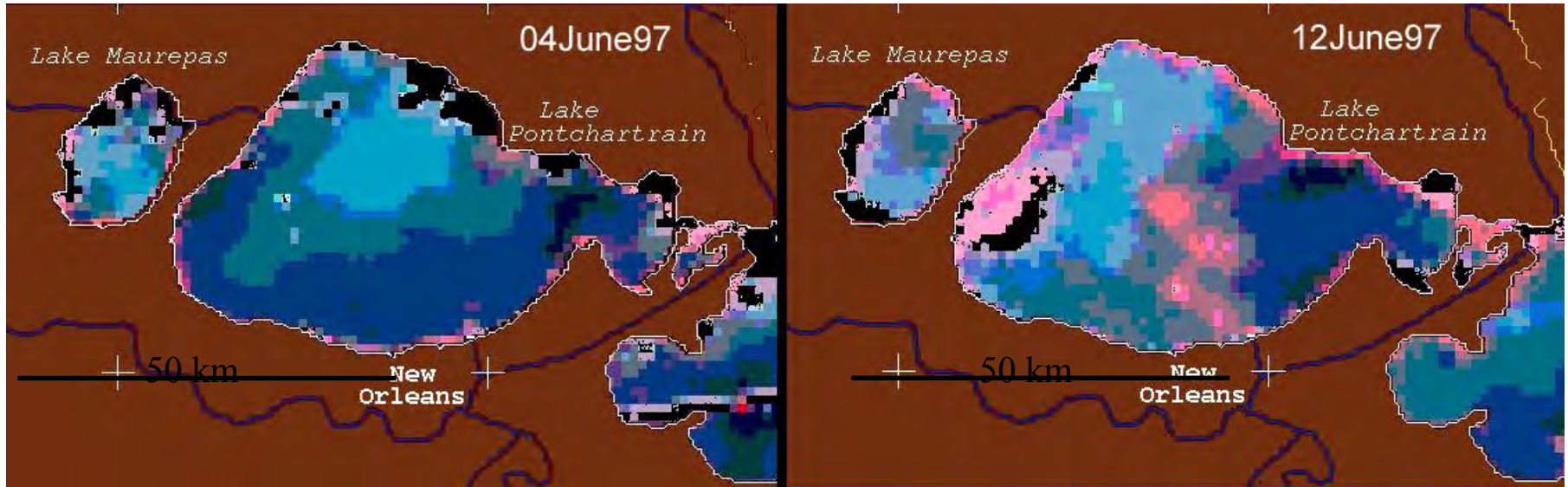
Lake Pontchartrain, Louisiana, USA

(Blue-green algae can produce surface scums)

False Color infrared (pink indicates high NIR reflectance)

Cell counts $< 10^3 \text{ mL}^{-1}$

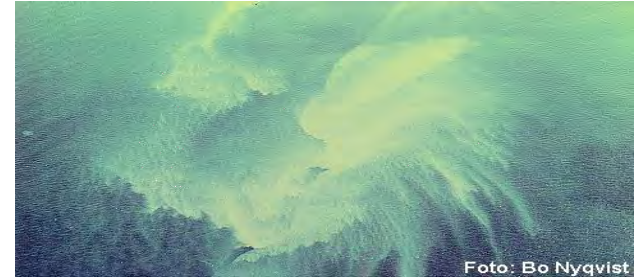
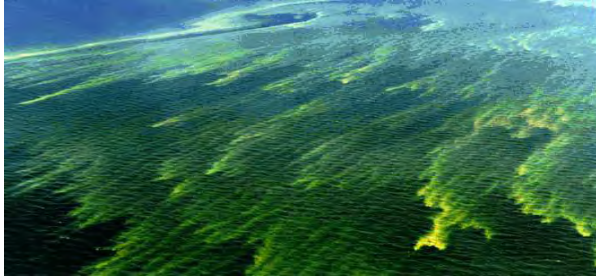
$> 10^5 \text{ mL}^{-1}$



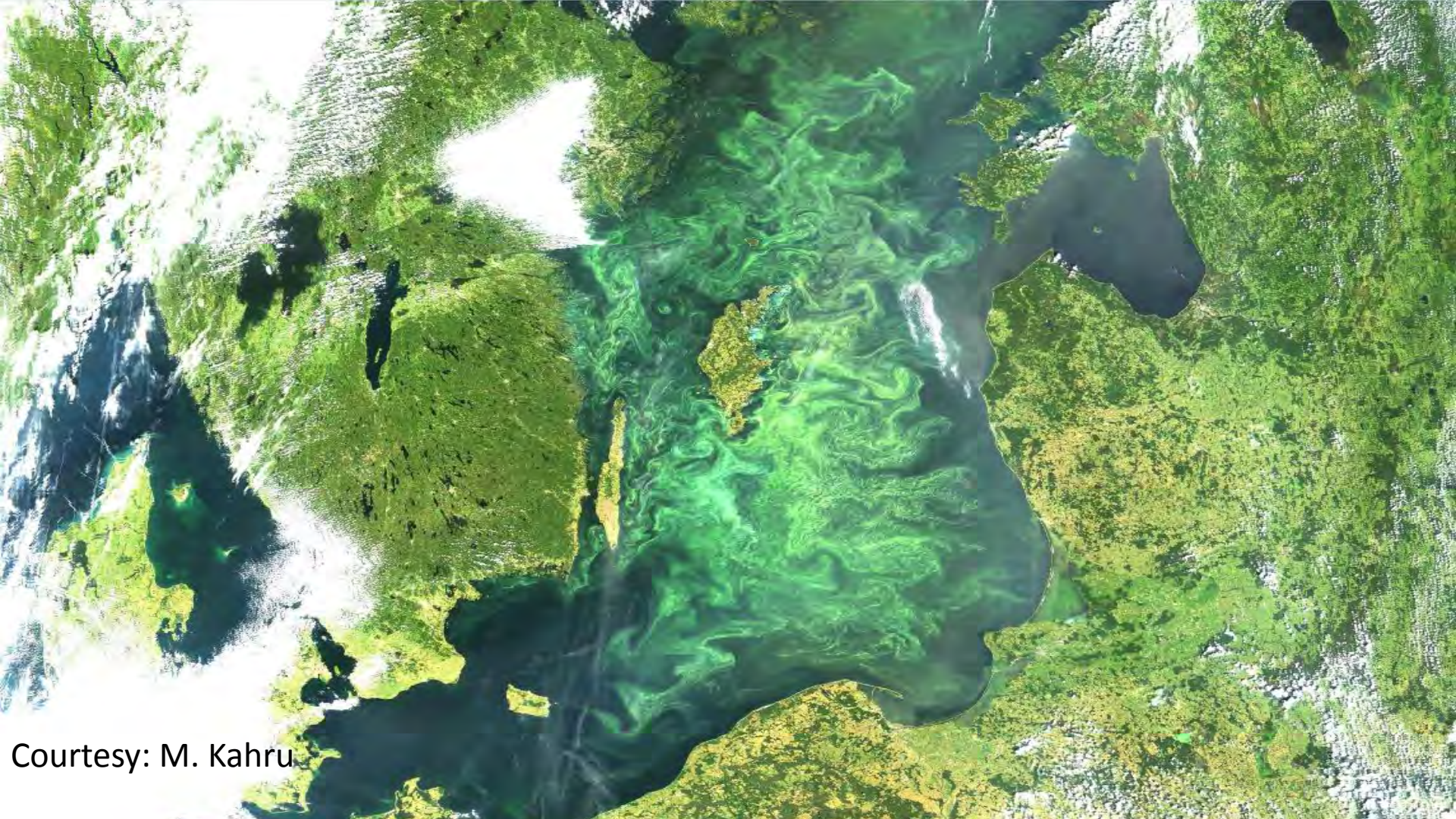
Extensive work done on blue-green blooms in the Baltic.

HABs with extreme scattering

(*Nodularia spumigena* dominated blooms in the Baltic Sea)



Strong scattering enhanced by the vacuoles in *Nodularia* cells (M. Kahru)



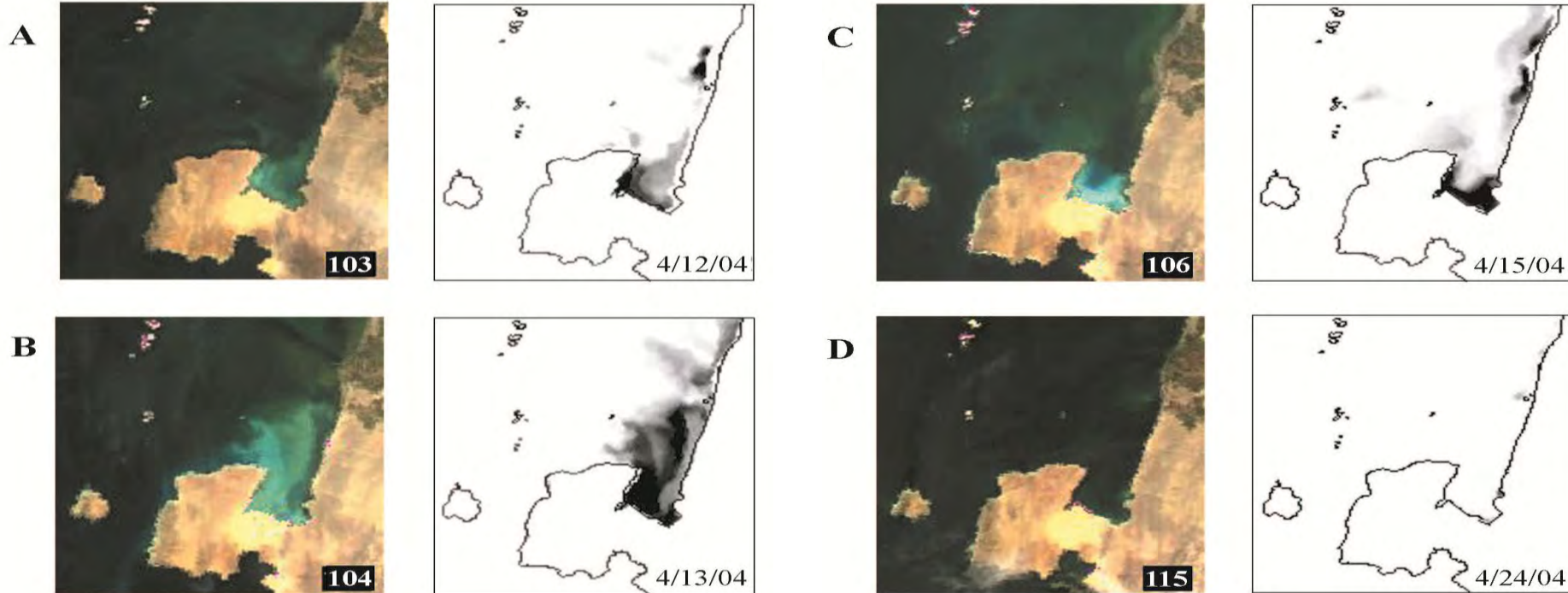
Courtesy: M. Kahru

Visible anomalies

Anoxic *Gymnodinium sanguineum* HAB in Peru, turbidity anomaly

True-color

“turbidity”



Kahru et al., Eos 2005

UV spectra, Mycosporine amino acids (MAAs) for dinoflagellates?

Sunscreen for algae

More common in dinoflagellates

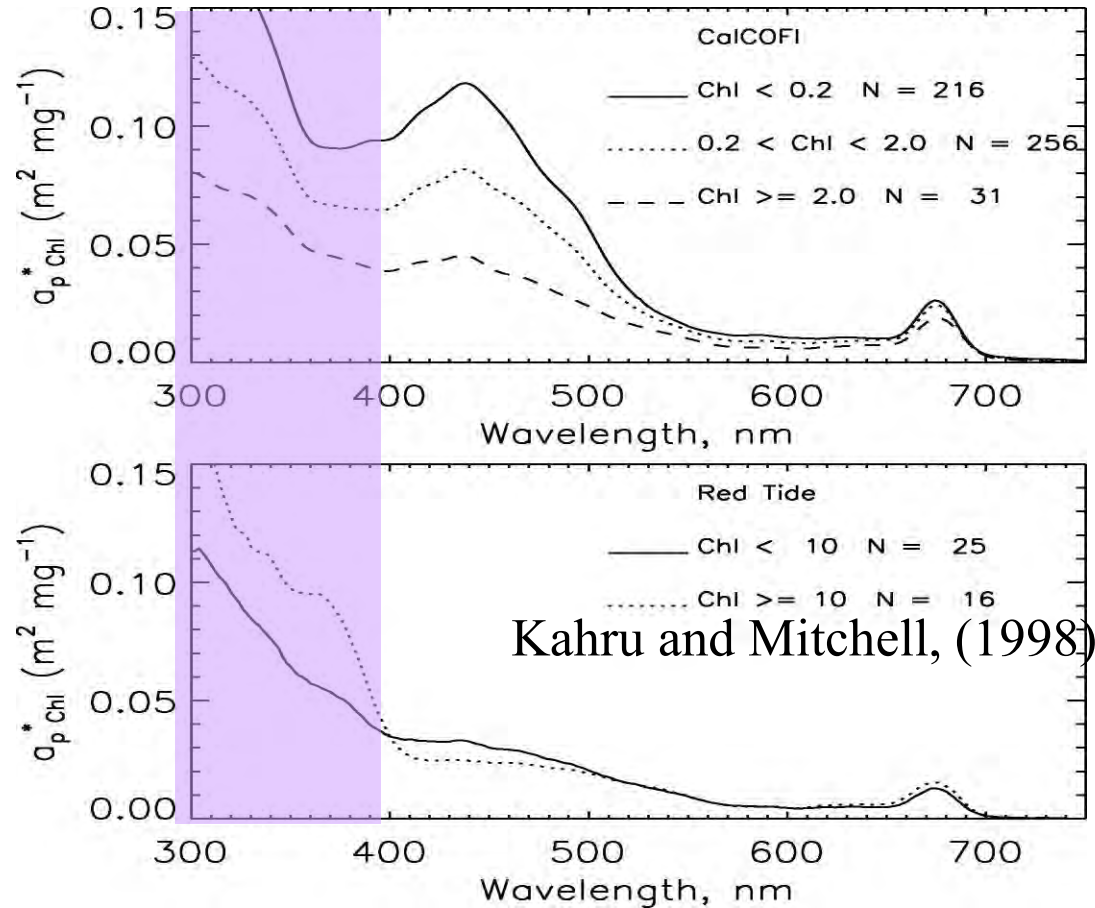
Increased UV absorption

Relevant with future UV sensors

Issues:

CDOM areas

Not stable, scene by scene classification



Common algorithms.

1. Chl-a blue:green
2. Chl-a NIR:red
3. FLH
4. Chlorophyll anomaly
5. MCI (SS-708)
6. CI (SS-681)
7. Bb/a ratio
8. Brightness
9. Multi-band empirical relationship

Exercise. Identify whether potential algal blooms occur. Feb and May images. CI, MCI, & OC4 chl

